Decomposing changes in phylogenetic and functional diversity over space and time

Loïc Chalmandrier1,2*, Tamara Münkemüller1,2, Vincent Devictor3, Sébastien Lavelgne1,2 and Wilfried Thuiller1,2

1Univ. Grenoble Alpes, LECA, F-38000 Grenoble, France; 2CNRS, LECA, F-38000 Grenoble, France; and 3Institut des Sciences de l’Évolution de Montpellier, UMR CNRS 5554, Université Montpellier 2, Place Eugène Bataillon, 34095 Montpellier Cedex 05, France

Summary
1. The α-, β-, γ-diversity decomposition methodology is commonly used to investigate changes in diversity over space or time but rarely conjointly. However, with the ever-increasing availability of large-scale biodiversity monitoring data, there is a need for a sound methodology capable of simultaneously accounting for spatial and temporal changes in diversity.
2. Using the properties of Chao’s index, we adapted Rao’s framework of diversity decomposition between orthogonal dimensions to a multiplicative α-, β-, γ-decomposition of functional or phylogenetic diversity over space and time, thereby combining their respective properties. We also developed guidelines for interpreting both temporal and spatial β-diversities and their interaction.
3. We characterized the range of β-diversity estimates and their relationship to the nested decomposition of diversity. Using simulations, we empirically demonstrated that temporal and spatial β-diversities are independent from each other and from α- and γ-diversities when the study design is balanced, but not otherwise. Furthermore, we showed that the interaction term between the temporal and the spatial β-diversities lacked such properties.
4. We illustrated our methodology with a case study of the spatio-temporal dynamics of functional diversity in bird assemblages in four regions of France. Based on these data, our method makes it possible to discriminate between regions experiencing different diversity changes in time. Our methodology may therefore be valuable for comparing diversity changes over space and time using large-scale data sets of repeated surveys.

Key-words: β-diversity, biodiversity, phylogenetic entropy, Shannon entropy, Hill numbers, diversity partitioning, bird assemblages, large-scale monitoring, turn-over

Introduction
Patterns of species diversity, as determined by their functional traits and phylogenetic relationships, have become central to addressing a large range of research questions such as the inference of assembly rules in community ecology (Diamond 1975; Webb 2000; Mouquet et al. 2012) or the delimitation of biodiversity hotspots in macro-ecology (Mazel et al. 2014). Using functional and phylogenetic diversity indices implicitly rejects the assumption that species are equally distinct entities, and instead accounts for their functional similarities and shared evolutionary history (Violle et al. 2007; Mouquet et al. 2012).

Thanks to the extension of large-scale biodiversity monitoring (Pereira & Cooper 2006) and the development of citizen science (Bonney et al. 2009), large data sets have been made available for investigating the spatial and temporal dynamics of biodiversity (Dornelas et al. 2014). These are of prime importance in evaluating how species assemblages are responding to ongoing changes in climate and land uses. Since these temporal changes are not necessarily homogeneous across space, a depiction of biodiversity changes from both a spatial and temporal perspective (Magurran et al. 2010) is required to understand which processes contribute to biodiversity dynamics. An adequate methodology is therefore needed to produce meaningful measures of diversity changes over space and time.

In his seminal paper, Whittaker (1960) proposed breaking the regional species diversity (γ-diversity) down into the average within-community species diversity (α-diversity) and the between community species diversity (β-diversity). More specifically, Whittaker formulated two laws to link α, β and γ-diversities: an additive law (γ = α + β) and a multiplicative law (γ = α × β). Two decomposition frameworks emerged from these two alternative approaches, each with different properties and drawbacks.

The additive law was adapted by Rao (1986) to the ‘Quadratic Entropy’ index which generalized the Gini–Simpson index to include species dissimilarities such as functional or phylogenetic distances. He further proposed decomposing γ-diversity into several dimensions (e.g. space and time), a procedure called Anodiv (Pavoine 2012). However, the additive decomposition of the γ-diversity, and by extension the Anodiv procedure, has been criticized for its inability to

*Correspondence author. E-mail: loic.chalmandrier@ujf-grenoble.fr

produce $\beta$-diversity estimates independent from the $\gamma$- and $\alpha$-diversities (Jost 2007; Basgela 2010). This property impedes Anodiv’s ability to access temporal or spatial biodiversity changes on large spatial scales. Indeed, large-scale biodiversity monitoring typically covers numerous regions with variable $\gamma$- and $\alpha$-diversities (e.g. Devictor et al. 2010), a consequence of the large-scale environmental filtering and historical contingencies that shape the biogeographical gradients of diversity (Hawkins, Porter & Diniz-Filho 2003). Should $\beta$-diversities be compared across regions, it is vital that they only quantify spatial or temporal change within these regions, independently from changes in $\gamma$ and $\alpha$-diversities. Otherwise these two effects would become indistinguishable.

The second framework, based on Whittaker’s multiplicative law, addresses this issue of independence. When diversity is calculated from an equivalent number (Hill 1973), it produces estimates of $\beta$-diversity which are independent from the $\alpha$- and $\gamma$-diversities (Jost 2007; Tuomisto 2010). Furthermore, the estimate of the $\beta$-diversity is set between 1 and the number of communities in the region studied. This property makes it possible to produce standardized estimates of $\beta$-diversity which are not dependent on the study design used in the region (Chao, Chiu & Hsieh 2012). This is a particularly important feature since large-scale biodiversity monitoring systems tend to be spatially and temporally beta diversities. Indeed, large-scale biodiversity monitoring typically covers numerous regions with variable $\gamma$- and $\alpha$-diversities (Chao, Chiu & Diniz-Filho 2003). Should they only quantify spatial or temporal change within these regions, independently from changes in $\gamma$ and $\alpha$-diversities. Otherwise these two effects would become indistinguishable.

We have built on these two frameworks and their respective advantages to propose a novel methodology for decomposing phylogenetic and functional diversity over space and time, and obtaining measurements of $\beta$-diversity which are independent of $\gamma$-diversity and $\alpha$-diversity. This study first introduces our multiplicative framework for estimating spatial and temporal beta diversities. Secondly, using a simulation-based approach, we demonstrate that in the case of taxonomic diversity, the estimated $\beta$-diversities are pairwise independent from the $\gamma$- and $\alpha$-diversity and from each other. Finally, we illustrate its novelty and features in a case study by decomposing the spatio-temporal effects on the functional diversity of the common avifauna in four regions of France over the last decade.

### Decomposing diversity over space and time

#### Definitions

We considered a region containing $S$ sites in which species were recorded at $T$ dates. We defined a community as the species composition of site $s$ at a given date $t$. We defined a site pool as the pool of all communities at site $s$ pooled for all dates, a time pool as the pool of all communities at a given date $t$ pooled for all sites and the regional pool as the pool of all communities for all sites and dates. The spatio-temporal decomposition of diversity (multiplicative $\alpha$, $\beta$, $\gamma$-decomposition) will ultimately be calculated based on the ratios of the diversities in these different units (communities and pools).

### Diversity Index

To calculate the diversity of a given unit, we used Shannon entropy exponential. This index is an ‘equivalent number’, part of the family of Hill numbers (Hill 1973). As such, its value ranges from 1 (if one species makes up most of the total abundance in the unit) to the number of species in the unit (if their relative abundances are all equal). It can be interpreted as the number of ‘equally abundant virtual species’ in the unit (Tuomisto 2010):

$$D(P) = \exp \left(- \sum_{i=1}^{N} \log(p_i) \times p_i \right) \quad \text{eqn 1}$$

with $P$, the vector $\{p_1, p_2, \ldots, p_N\}$ of abundances of the $N$ species present in the unit studied.

To include functional and phylogenetic similarities between species, we used the version of this index formulated by Chao, Chiu & Jost (2010) from Allen’s phylogenetic entropy (2009).

$$D(P) = \exp \left(- \sum_{b} \frac{L(b)}{T} \log(p_b(b)) \times p_b(b) \right) \quad \text{eqn 2}$$

where the summation is made over all branches of an ultrametric phylogenetic or functional tree of tips-to-root distance $T$, $L(b)$ is the length of branch $b$ and $p_b$ denotes the vector containing for each branch $b$, the summed relative abundance of its descendent species. We will refer to this index as Chao’s index.

Since the index includes species similarities, its absolute value can be interpreted as the number of ‘equally abundant and fully distinct virtual species’ in the study unit.

### Spatio-temporal Decomposition

We drew inspiration from the Anodiv procedure (Rao 1986; Pavoine 2012) to decompose diversity according to two orthogonal factors, here time and space, according to a multiplicative framework using Chao’s index. It is expressed as follows:

$$D(P_) = \frac{D(P_{..})}{\exp \left[ \sum_{i=1}^{T} \omega_i \log(D(P_{,i})) \right]} \times \frac{D(P_{..})}{\exp \left[ \sum_{s=1}^{S} \omega_s \log(D(P_{,s})) \right]} \times $$

$$\times \exp \left[ \sum_{i=1}^{T} \omega_i \log(D(P_{,i})) \right] \times \exp \left[ \sum_{s=1}^{S} \omega_s \log(D(P_{,s})) \right]$$

$$D(P_{,..}) \times \exp \left[ \sum_{s=1}^{S} \omega_s \log(D(P_{,s})) \right]$$

where $P_{,..}$ is the vector of species relative abundance in community at site $s$ and date $t$. The formulation includes $\omega_{si}$, a weight
attributed to a community at site s and time t, that sums to 1 over all s and t based for instance on total abundance or species richness in the community. A perfectly balanced design will involve the absence of missing data and the equal weighting of all communities, that is for all s and t, \( o_{st} = \frac{1}{N} \). Unequal weighting is not compulsory but is typically relevant when communities have been sampled with different sampling efforts (for a discussion, see Hardy & Jost 2008).

\( o_s \) and \( o_t \) are the weights of site pools s and time pools t, respectively, and are calculated as the sum of the weights of their constituent communities. The vector of species relative abundance for the site pool s and time pool t is thus calculated as the weighted mean of the species relative abundances in their constituent communities: \( P_s = \frac{1}{N_s} \sum_{i=1}^{S} o_{si} P_{si} \) and \( P_t = \frac{1}{N_t} \sum_{i=1}^{T} o_{ti} P_{ti} \). Finally, the vector of species relative abundances in the species pool is calculated as the weighted mean of species relative abundance across all communities: \( P_c = \sum_{s=1}^{S} \sum_{t=1}^{T} o_{st} P_{st} \).

Equation 3 can be reformulated as:

\[
\gamma = \frac{\gamma}{\beta_T} \times \frac{\gamma}{\beta_S} \times \frac{\alpha_S}{\alpha} \times \frac{\alpha_T}{\alpha} \times \frac{\alpha}{\alpha}
\]

with \( \gamma \) being the \( \gamma \)-diversity of the study region; with \( \alpha_T \) and \( \alpha_S \) being, respectively, the mean \( \alpha \)-diversity of time and site pools and \( \alpha \) the mean \( \alpha \)-diversity of the communities in the study region.

Or more simply,

\[
\gamma = \beta_T \times \beta_S \times \beta_{ST} \times \alpha
\]

with \( \beta_T \) being the temporal \( \beta \)-diversity, \( \beta_S \) the spatial \( \beta \)-diversity and \( \beta_{ST} \) the interaction term between the temporal and the spatial \( \beta \)- diversities. If the spatial and temporal structure of the data set is ignored, the total \( \beta \)-diversity \( \beta \) of the region across time can be expressed as:

\[
\beta = \frac{\gamma}{\alpha} = \beta_T \times \beta_S \times \beta_{ST}
\]

PROPERTIES OF THE \( \beta \)-DIVERSITIES

Chiu, Jost & Chao (2014) demonstrated that Chao’s index obeyed the ‘replication principle’. This implies that \( \beta_T \) and \( \beta_S \) have several of the properties enumerated in Jost (2007) and Tuomisto (2010), which facilitate the interpretation of their numerical values:

1. \( \beta_T \) and \( \beta_S \) are pairwise independent from \( \gamma \) and from \( \alpha_T \) and \( \alpha_S \), respectively (Jost 2007; Baselga 2010). Using simulations, we demonstrate below that \( \beta_T \) and \( \beta_S \) are pairwise independent from each other and are both pairwise independent from \( \alpha \).

2. The values of \( \beta_S \) (resp. \( \beta_T \)) are intuitive and can be interpreted as ‘the number of virtual, fully dissimilar and equally abundant site pools (resp. time pools)’ in the study region.

3. The values of \( \beta_S \), \( \beta_T \), and \( \beta \) have a range that is only dependent on the weights of, respectively, the site pools, time pools and the communities:

\[
1 \leq \beta_T \leq N_T, \text{ with } N_T = \exp \left[ -\sum_{t=1}^{T} o_t \times \log(o_t) \right]
\]

\[
1 \leq \beta_S \leq N_S, \text{ with } N_S = \exp \left[ -\sum_{s=1}^{S} o_s \times \log(o_s) \right]
\]

\[
1 \leq \beta \leq N_{ST}, \text{ with } N_{ST} = \exp \left[ -\sum_{s=1}^{S} \sum_{t=1}^{T} o_{st} \times \log(o_{st}) \right]
\]

The minimum possible value of \( \beta_S \), \( \beta_T \) and \( \beta \) will be 1 if the site pools, time pools or communities, respectively, are identical. The maximum possible value of \( \beta_S \), \( \beta_T \) and \( \beta \) will be attained if the site pools, time pools or communities, respectively, do not share species or tree branches. \( N_S \), \( N_T \) and \( N_{ST} \) can be interpreted as the equivalent number of sites pools, times pools and communities, respectively. If all communities are weighted equally, they will be equal to \( S \), \( T \) and \( ST \), respectively.

In other words, our measurement of change in diversity over space (resp. over time) has a natural minimum and maximum. Thus the absolute values of \( \beta_S \) and \( \beta_T \) can be standardized by their minimum and maximum value to make their value independent from the number of sites and time periods studied (Chao, Chiu & Hsieh 2012): \( \delta = \frac{\beta - \beta_{min}}{\beta_{max} - \beta_{min}} \) and \( \delta = \frac{\beta - \beta_{min}}{\beta_{max} - \beta_{min}} \).

\( \beta_{ST} \) has a minimum value of 1 and a maximum value constrained by both the value of \( \beta_S \) and \( \beta_T \) (see Appendix S2 for the demonstration),

\[
1 \leq \beta_{ST} \leq \min \left( \frac{N_{ST}}{N_S} \times \frac{1}{\beta_T}, \frac{N_{ST}}{N_T} \times \frac{1}{\beta_S} \right)
\]

RELATIONSHIP TO THE NESTED DECOMPOSITION OF DIVERSITY

Another methodological choice that can be made when analysing a spatio-temporal data set is to consider that space and time are nested (e.g. Sobek et al. 2009). If space is considered as nested within time periods, a new measure \( \beta_{ST} \) can be formulated to characterize the mean spatial \( \beta \)-diversity of the region within time periods (Pavoine & Dolédec 2005; Tuomisto 2010).

The decomposition will then be expressed as:

\[
\gamma = \beta_T \times \beta_{ST} \times \beta_S \times \alpha
\]

meaning that

\[
\beta_{ST} = \frac{\alpha_T}{\alpha} = \beta_S \times \beta_{ST}
\]

Alternatively, if we consider time periods as nested within space:

\[ \gamma = \beta_S \times \beta_{T/S} \times \alpha \]  

Eqn 8

meaning that

\[ \beta_{S/T} = \frac{\alpha_S}{\alpha} = \beta_T \times \beta_{ST} \]

Thus, \( \beta_{S/T} \) and \( \beta_{T/S} \) are not strictly \( \beta \)-diversities because they are the ratio of two mean \( \alpha \)-diversities from different hierarchical levels, rather than the ratio of a unit’s diversity and the diversity of its subunits. However, we demonstrated (Appendix S1) that like \( \beta \)-diversities, they have fixed minimum and maximum values that are independent of the \( \gamma \)- and \( \alpha \)-diversities:  

\[ 1 \leq \beta_{S/T} \leq \frac{\alpha_S}{S^2} \] and \[ 1 \leq \beta_{T/S} \leq \frac{\alpha_T}{S^2} \]

INTERPRETATION OF \( \beta \)-DIVERSITIES

The different \( \beta \)-diversities can be interpreted on their own and in combination with each other (Fig. 1). \( \beta_T \) quantifies the change in diversity time pools, or in other words, the temporal change in regional diversity, irrespective of the spatial patterns of diversity. \( \beta_S \) quantifies the change in diversity between site pools, or in other words, the spatial change in diversity after averaging the temporal variability of communities. \( \beta_{ST} \) quantifies the interaction between spatial and temporal turnover, it can be used to quantify finer changes such as a rearrangement of species between sites in two dates which are not quantified by \( \beta_S \) and \( \beta_T \) (Fig. 1). The case where \( \beta_{ST} \) is equal to 1 indicates that there is an identical change of diversity across space and time between communities, for instance, if between two dates a species is introduced in all studied communities at equal relative abundance. On the other hand, a value of \( \beta_{ST} \) over 1 denotes a heterogeneity of change of communities over space and time that is not quantified by \( \beta_S \) and \( \beta_T \) because it averages out at larger spatial or temporal scales. It is interesting to note that it is possible to have a situation where \( \beta_S \) and \( \beta_T \) equal one while \( \beta_{ST} \) is higher than one. This is illustrated in Fig. 1 (\( \beta_S = \beta_T = 1 \) and \( \beta_{ST} > 1 \)) with an extreme case in which two communities fully inversed their composition between the two dates studied. A concrete example could be the mosaic theory of forest regeneration (Remmert 1991): disturbances in a forested landscape would trigger the same temporal successions but at different times and locations generating a heterogeneous landscape. In this case, there is a spatial and temporal change between communities, but the time pools remain constant (in other words, the diversity of the region changes very little between the two dates) and the site pools also remain constant (in other words, when averaged over time, communities across the landscape have a similar composition).

TEST OF FUNCTIONAL AND PHYLOGENETIC \( \beta \)-DIVERSITIES

A common way to test the values of functional or phylogenetic \( \beta \)-diversities is to use a randomization model to generate a distribution of \( \beta \)-diversities under a certain null hypothesis. Our framework is compatible with any kind of randomization procedure. In the following case study of French avifauna, we chose the species shuffling procedure that has been shown to be among the most efficient null models in terms of Type I error rate (Hardy 2008). A significant high (resp. low) \( \beta \)-diversity thus indicates that species tend to be replaced by dissimilar (resp. similar) species over time or space. We calculated the effect size of each functional \( \beta \)-diversity, as the observed \( \beta \)-diversity minus the mean of its null distribution divided by the standard deviation of the null distribution. If the \( \beta \)-diversity was higher than expected (positive effect size), then the communities differed more than expected under a random assembly model; if the \( \beta \)-diversity was lower than expected (negative effect size), then the communities differed less than expected under a random assembly model.

Independence properties of the spatio-temporal diversity decomposition

We adapted the simulation procedures developed by Baselga (2010) to demonstrate the independence properties of our diversity decomposition framework. We used two simulation approaches: (i) a top-down approach where we first chose a \( \gamma \) value, then generated a community weight vector and sequentially randomly selected the values of \( \alpha_T, \alpha_S \) and \( \alpha \) and (ii) a bottom-up approach where we first chose an \( \alpha \) value, then generated a community weight vector and sequentially randomly selected the values of \( \alpha_T, \alpha_S \) and \( \gamma \). Each draw was constrained by minimum and maximal values deduced from the properties of the spatio-temporal decomposition stated above. For a given number of sites (S) and dates (T), we tested 200 initial \( \gamma \) or \( \alpha \) values between 1 and 200, each repeated 200 times. Both the top-down and bottom-up approach procedures are detailed in Appendix S3. Both procedures are necessary to demonstrate the pairwise independence of the \( \beta \)-diversities from \( \gamma \) and \( \alpha \) (Baselga 2010). In the top-down approach, the \( \beta \)-diversities need to be uncorrelated with \( \gamma \), and in the bottom-up approach, the \( \beta \)-diversities need to be uncorrelated with \( \alpha \).

When no data were missing (i.e. all sites observed at all times) and \( T = 4 \) and \( S = 10 \), we found no correlation of \( \gamma \) with \( \text{Std} \beta_T \) (Fig. 2, \( r = -0.0014 \); 95% CI interval: \([-0.011, 0.0084]\)) or \( \text{Std} \beta_S \) (Fig. 2, \( r = -0.0013 \); 95% CI interval: \([-0.011, 0.0085]\)) or \( \text{Std} \beta_T \) with \( \text{Std} \beta_S \) (Fig. 2, \( r = 0.0019 \); 95% CI interval: \([-0.012, 0.008] \)) in the top-down approach and no correlation of \( \alpha \) with \( \text{Std} \beta_T \) or \( \alpha \) with \( \text{Std} \beta_S \) in the bottom-up approach (Fig. S1. \( \alpha \) and \( \text{Std} \beta_T, r = 0.0052 \); 95% CI interval: \([-0.0046, 0.015] \)). In contrast, \( \beta_{ST} \) depended on the previously established \( \beta_T \) and \( \beta_S \) values, but is independent from both \( \gamma \) and \( \alpha \) (Fig. 2. \( \beta_{ST} \) and \( \gamma, r = 0.00038 \); 95% CI interval: \([-0.0094, 0.0010] \) Fig. S1. \( \beta_{ST} \) and \( \alpha, r = 0.0037 \); 95% CI interval: \([-0.0061, 0.0135] \)). When we further explored alternative \( T \) and \( S \) parameterizations (for any values of \( S \) and \( T \) between 2 and 10), we found these results to be robust (Table 1).

We also investigated the specific case where community data were missing in the data set. For all \( T \) and \( S \) parameter values, a varying proportion of community weights were set to 0 while
maintaining at least one community per site and date. We then studied how the amount of missing data affects the independence properties between $\gamma$, $\beta_T$, $\beta_S$, $\beta_{ST}$ and $\alpha$. We found that in the extreme case of perfect balance (all community weights are equal), all correlations remained close to 0. When we introduced unequal weighting and increased the proportion of missing data, the correlation between $\beta_T$ and $\beta_S$ increased slightly but remained on average close to 0. However, there were some extreme correlation values that deviated strongly from 0. We observed the same pattern for the other relationships although they tended to show more robustness (Fig. 3). This can be explained by the fact that the sampling design becomes less orthogonal between sites and dates as the amount of missing data increases. The extreme case would be a spatial-temporal data set where a single community was sampled per date, each time at different sites. Then $\beta_T$ and $\beta_S$ would be equal. However, as indicated by the correlation values, which were on average close to 0, the independence relationship remained on average quite robust and only a few weight vectors resulted in a loss of the independence properties.

Overall, we therefore conclude that $\beta_T$, $\beta_S$ and $\beta_{ST}$ are pairwise independent from $\alpha$ and $\gamma$. We further conclude that $\beta_T$ and $\beta_S$ are pairwise independent from each other but only when the weighting scheme does not deviate too far from a perfectly balanced sampling design (i.e. no missing data and for all $s$ and $t$, $\omega_{st} = \frac{1}{N}$).

Our simulation procedure has its limitations. Indeed, it is not clear how the inclusion of a phylogenetic or functional tree between species could further constrain the distribution of the different diversity metrics compared to the maximal values of

Fig. 1. Graphical interpretation of the eight possible patterns of diversity change over space and/or time and their respective $\beta_S$, $\beta_T$, $\beta_{ST}$ value. Each cell of the table represents one of the patterns illustrated by the composition of four communities from two sites (lines) at two dates (column), each represented by a circle. Sites and time pools (see Methods) composed from the pooled composition of communities over sites or dates, respectively, are represented by rectangles. Within communities and pools, symbols represent individuals from a species specified by the geometrical shape.

Case study: spatio-temporal changes in functional diversity in French bird assemblages

DATA

We applied our spatio-temporal framework to the avifauna monitored by the French breeding bird survey programme (Julliard et al. 2006). This programme relied on skilled ornithologists to monitor common birds using a standardized protocol from 2001 to 2012. Under this scheme, ornithologists recorded every individual seen or heard during a five-minute period at 10 count points, evenly distributed within 2 × 2 km survey sites. The sites were randomly selected around the observer’s locality, thus ensuring that a variety of habitats were monitored (including intensive farmlands, forests, suburbs and cities). We selected four regions, each defined as a circular window 100 km in diameter, belonging to two disparate biogeographical regions. Two were situated on the Mediterranean coast (MEDI and MED2) and two on the Atlantic coast (ATL1 and ATL2). Each region included different numbers of survey sites for which temporal trends were available (i.e. with sites monitored twice at least five years apart). ‘Communities’ were defined as the species assemblages recorded at the sites in the different regions. Species similarity was estimated from the ultrametric functional tree taken from Thuiller et al. (2014) based on body mass, diet and feeding behaviour (see Appendix S4 for details). Each community was given the same weight.

<table>
<thead>
<tr>
<th>Diversity measures</th>
<th>Top-down approach</th>
<th>Bottom-up approach</th>
</tr>
</thead>
<tbody>
<tr>
<td>γ and β_{TR}</td>
<td>[−0.0143, 0.0117]</td>
<td>[0.149, 0.598]</td>
</tr>
<tr>
<td>γ and β_{PS}</td>
<td>[−0.0137, 0.0153]</td>
<td>[0.146, 0.596]</td>
</tr>
<tr>
<td>γ and β_{ST}</td>
<td>[−0.0137, 0.0142]</td>
<td>[−0.181, −0.0653]</td>
</tr>
<tr>
<td>γ and α</td>
<td>[0.485, 0.912]</td>
<td>[0.485, 0.915]</td>
</tr>
<tr>
<td>S0_{γ} and β_{TR}</td>
<td>[−0.0137, 0.0270]</td>
<td>[−0.107, 0.193]</td>
</tr>
<tr>
<td>S0_{γ} and β_{PS}</td>
<td>[−0.559, −0.0725]</td>
<td>[−0.414, −0.185]</td>
</tr>
<tr>
<td>S0_{γ} and β_{ST}</td>
<td>[−0.486, −0.111]</td>
<td>[−0.062, 0.0136]</td>
</tr>
<tr>
<td>S0_{γ} and α</td>
<td>[−0.560, −0.0695]</td>
<td>[−0.415, −0.188]</td>
</tr>
<tr>
<td>S0_{β_{PS}} and β_{ST}</td>
<td>[−0.490, −0.106]</td>
<td>[−0.0145, 0.0124]</td>
</tr>
<tr>
<td>S0_{β_{PS}} and α</td>
<td>[−0.182, 0.0206]</td>
<td>[−0.0150, 0.0159]</td>
</tr>
<tr>
<td>β_{ST} and α</td>
<td>[−0.182, 0.0206]</td>
<td>[−0.0150, 0.0159]</td>
</tr>
</tbody>
</table>

environmental heterogeneity in MED1 and ATL2 compared to MED2 and ALT1. The null model made it possible to determine that the site pools of the region MED1 were more functionally different (albeit marginally) than expected from their taxonomic composition compared to ATL2 despite having a lower standardized value of spatial β-diversity (MED1, effect size of β5 = 1.22; ATL2, effect size of βT = 0.21; Fig. 4). The multivariate analysis of MED1 (Fig. S2) further suggested that sites at both dates were differentiated mainly according to the body size of their constituent species, suggesting a degree of large-scale environmental filtering acting on the birds’ functional traits. On the other hand, the difference between the ATL2 site pools was not significantly different from the random expectation. This suggests that the large functional difference between site pools was due to a large number of different species (hence the high spatial β-diversity) but that between the site pools, these species were not particularly distinct in terms of their functional traits.

Mathematically, this means that for all s and t, \( \omega_s = \frac{1}{T} \). To further facilitate the interpretation of the result of the spatio-temporal decomposition, we used a double principal component analysis (dpcoa, Pavoine, Dufour & Chessel 2004) to visualize the differences between the sites in each region in terms of functional composition (Appendix S5 and Fig. S2). A summary of the information on the four regions is available in Table 2.

Results

**Spatio-temporal diversity decomposition**

Overall, the temporal change in regional diversity was more substantial in the two Mediterranean regions than in the two Atlantic regions (Fig. 4). However, the use of the null model showed that the temporal β-diversity in MED1 was much higher than expected from the taxonomic change (effect size = 3.09) while in MED2, the temporal β-diversity was only marginally different from the taxonomic change (effect size = 1.48). This showed that between the two dates studied, the MED1 time pools were significantly different in terms of the functional traits of their constituent species (as suggested by the multivariate analysis, this was most likely due to the relative increase in larger-bodied species in all the sites over time, Fig. S2). In contrast, the two Atlantic regions showed low temporal β-diversity, and the null model further showed that it was even lower than expected from the taxonomic change for ATL1 (ATL1, effect size of βT = −1.87; ATL2, effect size of βT = 0.15; Fig. 4). This indicated that the sites studied in ATL1 remained remarkably constant over time, with species being substituted by other species with similar functional traits. Overall, these results are consistent with previous diachronic analyses, which demonstrated substantial changes in bird communities in inland Mediterranean areas over time due to major changes in land use (Preiss, Martin & Debussche 1997; Sirami, Brotons & Martin 2007).

**INTERACTION TERM**

The interaction term \( \beta_{ST} \) was always higher than 1 across the four regions studied. However, as the value of \( \beta_{ST} \) is dependent on both \( \beta_T \) and \( \beta_S \) and the study design (number of sites and number of dates), it was difficult to compare it across regions. The null model provides a way of determining whether the interaction term was higher or lower than expected from the taxonomic turnover. We found that the interaction term of \( \beta_{ST} \) was much higher than expected (effect size = 1.99) indicating that individual sites changed more in functional composition.
Furthermore, the case of ATL1 is interesting because while its effect size is low, its spatio-dynamic component is high. Firstly, it allows a standardized decomposition of a region with communities whose functional composition had changed markedly over time, but in opposite directions. This indicates potential for investigating community change across regions without inter-ference from the biogeographical differences quantified by the γ-diversities. In contrast, there was a risk with the original Anova procedure (Rao 1986) of yielding overestimated values for β-diversities that were unrelated to the number of sites or dates studied within each region (Chao, Chiu & Hsieh 2012). Although our method does not prevent bias if the sampling is not representative of the biodiversity in a region over space and time, the standardization makes it possible to compensate for an unbalanced study design between regions. In our case study, spatial and temporal functional β-diversities were standardized using the maximum and minimum value they could possibly attain in a region (Fig. 4), thus producing estimates of β-diversities that were unrelated to the number of sites or dates studied within each region (Chao, Chiu & Hsieh 2012). Although our method does not prevent bias if the sampling is not representative of the biodiversity in a region over space and time, the standardization makes it possible to compensate for an unbalanced study design between regions. This solves a common problem arising from large-scale biodiversity monitoring where remote areas tend to be under sampled (Jiguet et al. 2012; Ficetola et al. 2013).

Thirdly, our approach also makes it possible to test the space-for-time substitution often used when time-series data are not available. The drivers of diversity change such as climate (e.g. Blois et al. 2013) or land use (e.g. Sirami, Brotons & Martin 2007) can be studied both across space and time. Ecologists have thus traditionally used space-for-time substitution as an alternative to expensive and rare long-term studies (Pickett 1989; e.g. Chalmandrier et al. 2013). This substitution assumes that changes in diversity over spatial locations and changes in diversity over time are equivalent and independent under the assumption that they are driven by the same ecological process (Fukami & Wardle 2005). However, this assumption can easily be violated by confounding processes such as dispersal (Brotons, Pons & Herrando 2005), biotic interactions (Thuiller et al. 2007) and delayed responses to changes in the local environment (Devictor et al. 2012). Our methodology provides tools which are adapted to testing the assumption on the potential for investigating βST as an emergent spatio-dynamic component of community changes.

### Perspectives and limitations of spatio-temporal decomposition

The novel methodology presented herein has several advantages and strengths. Firstly, it allows a standardized decomposition of diversity across regions with potentially very different γ-diversities. In France, the functional and phylogenetic γ-diversities of avifauna are heterogeneous across space due to different macro-climatic influences (Devictor et al. 2010). Most notably, the Mediterranean Basin has high taxonomic, functional and phylogenetic γ-diversity, which contrasts with the rest of France. Using our methodology, β-diversities are found to be pairwise independent from the local γ-diversities, thus making it possible to study processes such as landscape heterogeneity or land use change across regions without interference from the biogeographical differences quantified by the γ-diversities. In contrast, there was a risk with the original Anova procedure (Rao 1986) of yielding overestimated values for β-diversities in biogeographical areas with high γ-diversities, regardless of the actual spatial or temporal change (Baselga et al. 2010).

Secondly, our methodology accounts for differences in sampling efforts between regions. In our case study, spatial and temporal functional β-diversities were standardized using the maximum and minimum value they could possibly attain in a region (Fig. 4), thus producing estimates of β-diversities that were unrelated to the number of sites or dates studied within each region (Chao, Chiu & Hsieh 2012). Although our method does not prevent bias if the sampling is not representative of the biodiversity in a region over space and time, the standardization makes it possible to compensate for an unbalanced study design between regions. This solves a common problem arising from large-scale biodiversity monitoring where remote areas tend to be under sampled (Jiguet et al. 2012; Ficetola et al. 2013).

### Table 2. Characteristics of the different regions

<table>
<thead>
<tr>
<th>Region</th>
<th>Biogeographical zone</th>
<th>Number of sites</th>
<th>Year</th>
<th>Year</th>
<th>Size of the species pool</th>
<th>Functional γ-diversity</th>
<th>Functional α-diversity</th>
</tr>
</thead>
<tbody>
<tr>
<td>MED1</td>
<td>Mediterranean</td>
<td>5</td>
<td>2003</td>
<td>2008</td>
<td>63</td>
<td>8.06</td>
<td>6.48</td>
</tr>
<tr>
<td>MED2</td>
<td>Mediterranean</td>
<td>14</td>
<td>2003</td>
<td>2009</td>
<td>57</td>
<td>10.75</td>
<td>7.22</td>
</tr>
<tr>
<td>ATL1</td>
<td>Atlantic</td>
<td>7</td>
<td>2003</td>
<td>2009</td>
<td>43</td>
<td>8.32</td>
<td>6.73</td>
</tr>
<tr>
<td>ATL2</td>
<td>Atlantic</td>
<td>5</td>
<td>2003</td>
<td>2009</td>
<td>44</td>
<td>8.69</td>
<td>6.10</td>
</tr>
</tbody>
</table>

which space-for-time substitution is based: the pairwise independence of $\beta_2$ and $\beta_3$ allows for the direct comparison of the spatial and temporal components of changes in diversity. However, we showed that this independence property was only maintained if the sampling design is balanced (i.e. there is relatively a low amount of missing data, and communities have a similar weight). We therefore recommend testing the sampling design beforehand using the simulation procedures to assess whether $\beta_2$ and $\beta_3$ are theoretically pairwise independent.

Fourthly, this is the first study to adapt the diversity decomposition between multiple factors originally proposed by Rao (1986) to the requirements of $\beta$-diversities computations, as recommended by Jost (2007) and Tuomisto (2010). Furthermore, we generalize this approach using Chao’s index (2009) which includes species’ functional and phylogenetic distances, thereby combining the advantages of both methods. Shannon entropy exponential and its generalization are the only equivalent numbers that fully combine the properties of the additive and multiplicative $\alpha$, $\beta$, $\gamma$-decomposition (Jost 2007). This opens promising avenues for adapting our framework to methodologies based on the additive decomposition of the Shannon entropy (e.g. Pélissier & Couteron 2007). It is also the only equivalent number where (i) there is more or less a general consensus about the handling of unequal weighting of communities (but see Chiu, Jost & Chao 2014) and (ii) the unequal weighting still leads to $\beta$-diversities values that are pairwise independent from $\gamma$- and $\alpha$-diversities (Jost 2007; Tuomisto 2010).

Chao’s index belongs to a large family of indices that extend the Hill numbers (1973) to include species’ phylogenetic similarities (Chao, Chiu & Jost 2010) making it possible to explicitly parameterize the weight given to a rare vs. a dominant species. While our framework is transposable to these indices, some properties (pairwise independence of $\beta$-diversities from $\gamma$- and $\alpha$-diversities, range of $\beta_{ST}$ and of nested $\beta$-diversities) need to be demonstrated, in particular in the case of missing data and unequal weighting of communities. The Chao’s index studied is based on assumptions on how to take into account species abundances, that is the contribution of a species to the diversity value is proportional to its relative abundance (Chiu, Jost & Chao 2014). It thus may not be suitable for achieving certain analytical aims: for instance, a conservation approach may want to consider rare and dominant species equally regardless of their relative abundance. On the other hand, a focus on ecosystem functioning may require an emphasis on dominant species as they are expected to be the main contributors to ecosystem functioning (Garnier et al. 2004; but see Mouillot et al. 2013). Furthermore, recent work has shown the value of analysing diversity patterns with multiple equivalent numbers in order to vary the weighting given to dominant as opposed to rare species and to disentangle multiple assembly rules (Arroyo-Rodriguez et al. 2013; Chalmandrier et al. 2014b). We therefore argue for more statistical development to adapt spatio-temporal decomposition to other equivalent numbers, using the generalization of Hill numbers, and thus adding a supplementary parameter which explicitly examines the impact of species’ relative abundances.

**Conclusion**

Recent years have seen major efforts to unify methodologies for evaluating and decomposing assemblage diversity. We have drawn on these achievements to propose a methodology that overcomes the challenges encountered when studying large-scale diversity data sets which encompass multiple orthogonal dimensions. We have shown that this approach can be used with classical animal survey data (also available for butterflies, fishes and plants) and that it provides clear results. Although more work is required to expand this method to multiple diversity indices, we believe that the properties of our methodology open up promising avenues for evaluating and testing diversity change across multiple dimensions. This will allow thorough analyses of the ever-increasing data produced by biodiversity survey programmes worldwide.

**Acknowledgements**

We warmly thank the hundreds of volunteers who took part in the French national breeding bird survey (STOC EPS programme). The research leading to these results received funding from the European Research Council under the European Community’s Seven Framework Programme FP7-2007-2013 Grant Agreement no. 281422 (TEEMBIO). TM was funded by the ANR-BiodivERSA project CONNECT (ANR-11-EBIB-002), as part of the ERA-Net BiodivERSA 2010 call. The LECA is part of LabEx OSUG@2020 (ANR10 LABX56).

**Data accessibility**

The site-by-species matrices and the functional tree of French avifauna are available online from the Dryad Digital Repository (Chalmandrier et al. 2014a).

**References**


